

1 **Xpert[®] MTB/RIF for the rapid diagnosis of tuberculous**
2 **lymphadenitis from Fine Needle Aspiration biopsy specimens**

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21 **Running Head:** Xpert[®] MTB/RIF on FNAB of lymph nodes

22 **Key words:** tuberculosis, FNAB, Xpert[®] MTB/RIF

23 **Conflicts of interest:** None to declare

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25 **Word Count Abstract: 51**

26 **Word Count Text: 1019**

27 **ABSTRACT**

28

29 This study demonstrates the excellent diagnostic accuracy of the Xpert[®] MTB/RIF test in
30 patients with tuberculous lymphadenitis. The test sensitivity and specificity was 96.7%
31 (95%CI, 86.6-100) and 88.9% (95%CI, 69.6-100) respectively, and correctly identified
32 6/6 (100%) of the cytology smear negative/culture positive cases and 1 of 2 (50%)
33 rifampin resistant cases.

34 **TEXT**

35 Tuberculous lymphadenitis is the most common extrapulmonary manifestation of TB
36 (4,8), and the majority of cases have no active lung involvement. Fine needle aspiration
37 biopsy (FNAB) offers a feasible and safe option for specimen collection (11,15). The use
38 of cytology together with the confirmation of acid-fastness by Ziehl-Neelsen staining and
39 Papanicolaou stain-induced fluorescent microscopy as well as mycobacterial detection by
40 culture offers excellent yields (13,14), but remains limited by the absence of species
41 confirmation, slow turn-around times and/or lack of drug resistance guidance.
42 Conventional microbiological culture and drug susceptibility testing are not always
43 available and in rare instances may take 6 weeks or longer (10).

44

45 The World Health Organization (WHO) endorsed Xpert[®] MTB/RIF combines sample
46 processing and real time polymerase chain reaction (PCR) in a fully automated platform
47 and detects *Mycobacterium tuberculosis* complex and rifampin resistance in less than 2
48 hours (2,3,9). Xpert[®] MTB/RIF has been used successfully on various extrapulmonary
49 specimens including urine and stool (6), but has not been rigorously evaluated with the
50 use of tissue specimens such as FNAB.

51

52 To determine the diagnostic utility of the Xpert[®] MTB/RIF, FNAB were collected from
53 50 consenting patients by aspirating two passes of a 23- or 25-g needle attached to a 10ml
54 syringe (IRB 05/03/043). Two smears were prepared from each aspirate, one fixed with
55 commercial cytology fixative for Papanicolaou staining and evaluation by fluorescent
56 microscopy and the other air dried for Giemsa and subsequent ZN staining. Smears were

57 evaluated for adequacy and for a morphological diagnosis and cases were excluded from
58 the analysis if either one or both passes had inadequate cellular material on smears. Both
59 ZN and Papanicolaou stain-induced fluorescent microscopy evaluations were performed
60 for direct detection of mycobacteria on all specimens. Residual material from one of the
61 aspirates was rinsed in a Mycobacterial Growth Indicator Tube (MGIT 960, Becton
62 Dickinson, USA) by aspirating a small volume of fluid into the syringe and expressing it
63 back into the MGIT 960 tube, followed by incubation in a MGIT 960 instrument for
64 mycobacterial culture. Positive cultures were identified as *Mycobacterium tuberculosis*
65 complex and genotypic drug susceptibility testing was done using Genotype MTBDR*plus*
66 assay (Hain Lifesciences, Germany) (1).

67

68 The residual material from the remaining aspirate was rinsed as above into 0.7 ml sterile
69 phosphate buffered saline (PBS) in a 10ml headspace glass vial sealed with a TFE/Sil
70 Septa and Aluminum open top seal. Sample preparation buffer was then added to the vial
71 in a 2:1 ratio, incubated at room temperature and subsequently processed for Xpert[®]
72 MTB/RIF testing as previously described (2).

73

74 Performance calculations, including test sensitivity, specificity and predictive values were
75 done using Statistica version 8 to compare the diagnostic performance of the Xpert[®]
76 MTB/RIF test to the reference standard (as positive cytology (cytomorphology consistent
77 with mycobacterial infection *and* direct visualization of the organism on ZN and/or
78 Papanicolaou stain-induced fluorescent microscopy) and/or culture positive for *M.*
79 *tuberculosis* (11,12)).

80

81 Of the 50 patients recruited, 48 cases had adequate smears for diagnosis (see Figure 1 for
82 patient recruitment flow diagram). In total, cytomorphological features associated with
83 TB were seen in 32 (66.7%) patients, non-specific reactive nodes identified in 10
84 (20.8%), acute bacterial lymphadenitis in 1 (2.1%), malignancy in 4 (8.3%) epithelial
85 inclusion cyst in 1 (2.1%) (Table 1).

86

87 Compared to the reference standard, Xpert[®] MTB/RIF correctly identified 29 out of 30
88 TB cases (sensitivity 96.7%, 95%CI, 86.6-100) (Table 2). The possible “false negative”
89 result had a prolonged transit interval of 9 days before Xpert[®] MTB/RIF testing, which
90 may have affected the result. Xpert[®] MTB/RIF was positive in two cases with negative
91 cytomorphology and culture (specificity 88.9%, 95%CI, 69.6-100). The cytomorphology
92 from one of the “false positive” results showed a necrotizing suppurative lymphadenitis,
93 which is consistent with TB. However, no organisms could be identified on microscopy
94 or culture. The cytomorphology of the other false positive result showed an epithelial
95 inclusion cyst, and the reason for this false positive result remains unknown. One case
96 had a negative culture result with positive cytology (including mycobacterial
97 identification) and a positive Xpert[®] MTB/RIF test. This patient had been on TB
98 treatment for one month at the time of specimen collection, which provides the likely
99 explanation for this discrepant result.

100

101 The Xpert[®] MTB/RIF test was positive in all 6 smear negative culture positive cases and
102 correctly identified the 1 of the 2 rifampin resistant cases. The average time to result for

103 microbiological culture was 18.5 days (range 9-55 days), while the Xpert[®] MTB/RIF test
104 result was available within 2 hours of commencing the test. This represents a substantial
105 reduction in diagnostic delay, thereby permitting real-time decision making and planning
106 of treatment (5).

107

108 A recent study by Hillemann *et al.* demonstrated the effectiveness of Xpert[®] MTB/RIF on
109 extrapulmonary tissue (6). In that study, the combined sensitivity and specificity of
110 77.3% and 98.2% was reported, respectively. Our study is the first to evaluate the
111 performance of Xpert[®] MTB/RIF in diagnosing tuberculous lymphadenitis through the
112 use of FNAB specimens. Study limitations include the small number of rifampin
113 resistant cases identified and the fact that the research was conducted in a referral center,
114 as ideally the technique is suited to use in peripheral laboratories to be effective in
115 controlling the disease. A positive aspect of the study is that the patient population is
116 representative of patients presenting with peripheral lymphadenopathy in most TB/HIV
117 endemic areas. It is unlikely that our patient cohort had exacerbated disease as compared
118 to patients presenting at primary health-care clinics as these patients are routinely referred
119 from the primary health-care clinic to the referral centre for FNAB.

120

121 In conclusion, FNAB is a simple procedure which can be performed in an outpatient
122 setting by clinicians or nursing staff after a short training period (7,15). It is ideal for use
123 in resource-limited settings, including more remote and rural areas (15). Specimen
124 collection is simple and safe. With the use of a transport vial virtually no sample
125 preparation is required and there is minimal risk of contamination. Furthermore, the

126 transmission risk to the operator may also be reduced. Combining FNAB and rapid
127 genotypic diagnosis using automated systems should greatly improve access to
128 appropriate diagnosis and treatment for patients with tuberculous lymphadenitis.
129

130 **ACKNOWLEDGMENTS**

131 We thank Mr. Justin Harvey (Stellenbosch University) for statistical assistance.

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189

190 **Table 1: Demographics and diagnostic outcome of patients referred for fine**
 191 **needle aspiration biopsy with possible mycobacterial lymphadenitis.**
 192

	Number (N=48)	Percent
Age		
<5 years	2	4.2
5-20 years	6	12.5
>20 years	40	83.3
Gender		
Male	20	41.7
Female	28	58.3
Patients on TB treatment	2	4.2
HIV infection status		
Positive	9	18.8
Negative	3	6.6
Unknown	36	75
Culture +/-HIV +	4/9	44.4
Cytological features		
Reactive lymph node	10	20.8
Features consistent with TB	32	66.7
Acute bacterial infection	1	2.1
Malignant or suggestive of malignancy	4	8.3
Epithelial inclusion cyst	1	2.1

Cases with cytomorphology suggestive of TB

Smear + ^a , Culture +, GeneXpert [®] +	22	45.8
Smear + ^a , Culture +, GeneXpert [®] -	1	2.1
Smear -, Culture +, GeneXpert [®] +	6	12.5
Smear + ^a , Culture -, GeneXpert [®] +	1	2.1
Smear -, Culture -, GeneXpert [®] +	1	2.1
Smear -, Culture -, GeneXpert [®] -	1	2.1

193

194 ^a Smear + = Cytomorphology suggestive of TB with direct visualization of organism.

195 + positive, - negative

196

197 **Table 2: Diagnostic accuracy of the Xpert[®] MTB/RIF test versus various reference**
 198 **standards.**
 199

Reference standard	Sensitivity		Specificity		Predictive values	
	n/N (%)	95% CI	n/N (%)	95% CI	PPV (%)	NPV (%)
Xpert [®] vs. Reference Standard*	29/30 (96.6)	86.6-100	16/18 (88.9)	69.6-100	93.5	94.1
Xpert [®] vs. Culture	28/29(96.6)	86.1-100	16/19(84.2)	63.6-100	90.3	94.1
Xpert [®] vs. smear- Culture+	6/6(100)	100	(100)	100	100	100
Autofluorescence vs. Culture	22/29(75.9)	60.3-91.4	18/19(94.7)	79.3-100	95.7	72
ZN vs. Culture	12/29(41.4)	23.5-59.3	19/19	100	100	52.8

200

201 *Reference standard as defined in the text – positive cytology (cytomorphology
 202 consistent with mycobacterial infection with direct visualization of the organism) and/or
 203 mycobacterial culture. ZN = Ziehl-Neelsen stain; n = index group; N = control group

204 **LEGENDS**

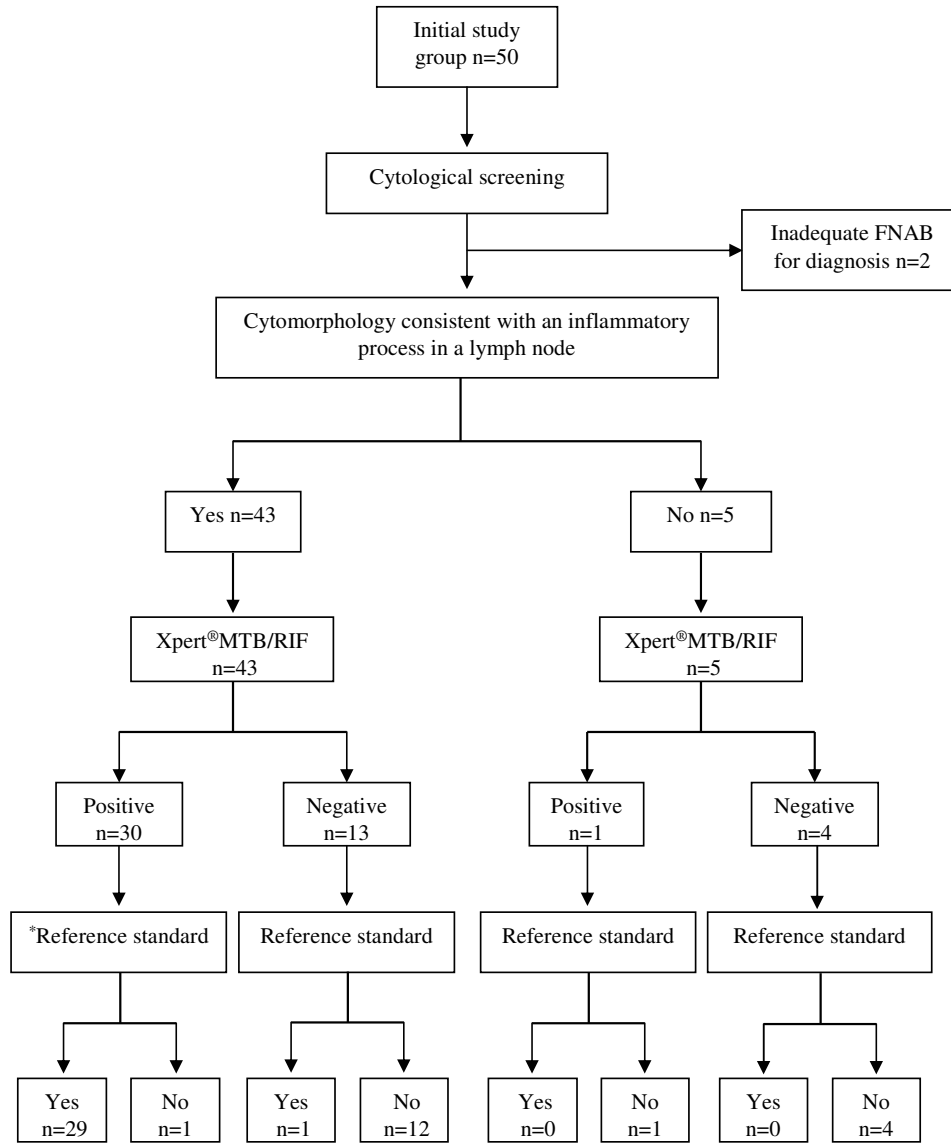
205

206 **Figure 1.** Flow diagram of all patients referred for fine needle aspiration biopsy with

207 possible mycobacterial lymphadenitis.

208

Figure 1



* Reference standard = Cytomorphology suggestive of TB with direct visualization of the organism and/or bacteriological culture
 FNAB = Fine Needle Aspiration Biopsy